## Dilution, degradation, and time delays in Boolean network models

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#### Motivation

We've seen how to incorporate the following features into ODE models:

- dilution of protein concentation due to cellular growth;
- degradation (or decay) of protein concentration;
- time-delays due to cellular processes.

In this section, we'll see how to add these types of features to Boolean models.

Our Boolean models will be derived from the 3-variable and 5-variable ODE models from the previous lecture.

## Dilution and degradation

Suppose Y regulates the production of X.

Assume Y(t) = 1 implies X(t + 1) = 1. (activation takes 1 step).

Generally, the loss of X due to dilution and degradation takes several steps.

Introduce new variables  $X_{\mathsf{old}(1)}, X_{\mathsf{old}(2)}, \ldots, X_{\mathsf{old}(n)}$ .

### **Properties**

- (i) If Y(t) = 0 and X(t) = 1, then  $X_{old(1)}(t+1) = 1$ . ("X has been reduced once by dilution & degradation.")
- (ii) If Y(t)=0 and  $X_{\mathrm{old}(i-1)}(t)=1$ , then  $X_{\mathrm{old}(i)}(t+1)=1$ . ("X has been reduced i times by dilution & degradation.")
- (iii) The number of "old" variables is determined by the number of timesteps required to reduce [X] below the discretation threshold.

Thus, X(t + 1) = 1 when either of the following holds:

- Y(t) = 1 (new amount will be produced by t + 1),
- $X(t) \wedge \overline{X_{\text{old}(n)}(t)} = 1$  (previous amounts of X still available).

$$X(t+1) = Y(t) \vee \left(X(t) \wedge \overline{X_{\mathsf{old}(n)}(t)}\right)$$

#### Other features

### Time delays

Say R regulates production of X, delayed by time  $\tau$  (n steps).

Introduce new variables  $R_1, R_2, \ldots, R_n$ , with transition functions:

$$R_{1}(t+1) = R(t)$$

$$R_{2}(t+1) = R_{1}(t)$$

$$R_{3}(t+1) = R_{2}(t)$$

$$\vdots$$

$$R_{n}(t+1) = R_{n-1}(t)$$

$$X(t+1) = R_{n}(t)$$

#### Medium levels of lactose

Introduce a new variable  $L_m$  meaning "at least medium levels" of lactose. Clearly, L=1 implies  $L_m=1$ .

- High lactose: L = 1,  $L_m = 1$ .
- Medium lactose: L = 0,  $L_m = 1$ .
- Basal lactose levels: L = 0,  $L_m = 0$ .

We can ignore any state for which L = 1,  $L_m = 0$ .

## Estimating constants for our Boolean model

## 3-variable ODE model of the lac operon (Yildirim and Mackey, 2004)

Let M(t)= mRNA,  $B(t)=\beta$ -galactosidase, and A(t)= allolactose (concentrations), respectively.

$$\begin{split} \frac{dM}{dt} &= \alpha_M \frac{1 + K_1 (e^{-\mu \tau_M} A_{\tau_M})^n}{K + K_1 (e^{-\mu \tau_M} A_{\tau_M})^n} - \widetilde{\gamma_M} M \\ \frac{dB}{dt} &= \alpha_B e^{-\mu \tau_B} M_{\tau_B} - \widetilde{\gamma_B} B \\ \frac{dA}{dt} &= \alpha_A B \frac{L}{K_L + L} - \beta_A B \frac{A}{K_A + A} - \widetilde{\gamma_A} A \end{split}$$

We need to estimate these rate constants and time delays from the literature.

- Time delays:  $\tau_M = .10 \text{ min}$ ,  $\tau_B = 2.00 \text{ min}$ .
- Degradtion rates are harder to determine experimentally, and they vary widely in the literaure. Sample values:

$$\left\{ \begin{array}{l} \gamma_A = .52 \; \mathrm{min}^{-1}, \quad .0135 \; \mathrm{min}^{-1}, \quad .00018 \; \mathrm{min}^{-1} \\ \gamma_B = .00083 \; \mathrm{min}^{-1}, \\ \gamma_M = .411 \; \mathrm{min}^{-1}, \\ \mu \in (.0045, \; .0347) \end{array} \right.$$

# Estimating constants for our Boolean model

### Approach

We'll select "middle of range" estimates for the rate constants:

$$\mu = .03 \text{ min}^{-1}$$
,

$$\mathbf{P}$$
  $\gamma_A = .014 \ \mathrm{min}^{-1}$   $\Longrightarrow$   $\widetilde{\gamma_A} = \gamma_A + \mu = .044$ ,

$$\blacksquare \ \gamma_B = .001 \ \mathrm{min}^{-1} \qquad \Longrightarrow \qquad \widetilde{\gamma_B} = \gamma_B + \mu = .031,$$

$$\mathbf{P}$$
  $\gamma_M = .411 \, \mathrm{min}^{-1} \qquad \Longrightarrow \qquad \widetilde{\gamma_M} = \gamma_M + \mu = .441.$ 

Degradation is assumed to be exponential decay: x' = -kx implies  $x(t) = Ce^{-kt}$ .

The half-life is the time t such that:

$$x(t) = Ce^{-kt} = .5C$$
  $\Longrightarrow$   $e^{-kt} = .5$   $\Longrightarrow$   $-kt = \ln \frac{1}{2}$   $\Longrightarrow$   $t = \frac{\ln 2}{k}$ 

#### Half-lives

$$\widetilde{h_A} = \frac{\ln 2}{\widetilde{\gamma_A}} = 15.753$$
 (approx. 1 time-step to decay)

$$\widetilde{h_B} = \frac{\ln 2}{\widetilde{\gamma_B}} = 22.360$$
 (approx. 2 time-steps to decay)

$$ightharpoons$$
  $\widetilde{h_M}=rac{\ln 2}{\widetilde{\gamma_M}}=1.5$  (approx. 0 time-steps to decay)

### Model assumptions

- Variables are M, B, A.
- Glucose absent. Intracellular lactose present, two parameters: L and  $L_m$ .
- Time-step ≈ 12 min.
- Ignore (all  $\ll$  12):  $\tau_M = .10$  min,  $\tau_B = 2$  min,  $\widetilde{h_M} = 1.572$  min.
- Introduce variables for dilution and degradation:
  - $A_{\text{old}}$  (since  $\widetilde{h_A} \approx 15.8 \approx 1 \text{ timestep}$ )
  - $B_{\text{old}}$ ,  $B_{\text{old}(2)}$  (since  $\widetilde{h_B} \approx 22.4 \approx 2$  timesteps)

### Proposed model

$$\begin{split} f_M &= A & f_B &= M \vee \left( B \wedge \overline{B_{\text{old}(2)}} \right) \\ f_A &= \left( B \wedge L_m \right) \vee L \vee \left( A \wedge \overline{A_{\text{old}}} \wedge \overline{B} \right) & f_{B_{\text{old}(1)}} &= \overline{M} \wedge B \\ f_{A_{\text{old}}} &= \left( (\overline{B} \vee \overline{L_m}) \wedge \overline{L} \right) \wedge A & f_{B_{\text{old}(2)}} &= \overline{M} \wedge B_{\text{old}(1)} \end{split}$$

Most of the functions should be self-explanatory.

### Justification for $f_A$

$$f_A = (B \wedge L_m) \vee L \vee (A \wedge \overline{A_{old}} \wedge \overline{B})$$

There are 3 ways for allolactose to be available at t + 1:

- (i)  $\beta$ -galactosidase and at least medium levels of lactose are present;
- (ii) high levels of lactose (assume basal concentrations of  $\beta$ -galactosidase);
- (iii) Enough allolactose is present so that it's not degraded below the threshold, and no  $\beta$ -galactosidase is present.

Let's write our model into polynomials form, with parameters  $(L,L_m)$  and variables  $(x_1,x_2,x_3,x_4,x_5,x_6)=(M,A,A_{\text{old}},B,B_{\text{old}(1)},B_{\text{old}(2)})$ :

$$\begin{array}{ll} f_M = A & f_1 = x_2 \\ f_A = (B \wedge L_m) \vee L \vee \left(A \wedge \overline{A_{\text{old}}} \wedge \overline{B}\right) & f_2 = x_2(1+x_3)(1+x_4) + (L_mx_4 + L + x_4LL_m) \\ & + x_2(1+x_3)(1+x_4) + (L_mx_4 + L + x_4LL_m) \\ f_{A_{\text{old}}} = \left((\overline{B} \vee \overline{L_m}) \wedge \overline{L}\right) \wedge A & f_3 = (1+x_4L_m)(1+L)x_2 \\ f_B = M \vee \left(B \wedge \overline{B_{\text{old}(2)}}\right) & f_4 = x_1 + x_4(1+x_6) + x_1x_4(1+x_6) \\ f_{B_{\text{old}(2)}} = \overline{M} \wedge B & f_5 = (1+x_1)x_4 \\ f_{B_{\text{old}(2)}} = \overline{M} \wedge B_{\text{old}(1)} & f_6 = (1+x_1)x_5 \end{array}$$

# Using Sage to compute the fixed points (high lactose)

```
1
2
P.<x1,x2,x3,x4,x5,x6> = PolynomialRing(GF(2), 6, order ='lex'); P
3
Multivariate Polynomial Ring in x1, x2, x3, x4, x5, x6 over Finite Field of size 2
4
L=1;
L=1;
L=1;
print "L =", L;
print "L h = ", Lh;

1
I = ideal(x1+x2, x2+(L*x4+Lh*x4*L*Lh)+(x2*(1+x3)*(1+x4))+(L*x4+Lh*x4*L*Lh)*(x2*(1+x3)*(1+x4)), x3+(1+x4*L)*(1+Lh)*x2, x4*x1*x4*(1+x6)*x1*x4*(1+x6), x5+(1+x1)*x4, x6+(1+x1)*x5); I
1
Ideal (x1 + x2, x2 + 1, x3, x1*x4*x6 + x1*x4 + x1 + x4*x6, x1*x4 + x4 + x5, x1*x5 + x6) of Multivar iate Polynomial Ring in x1, x2, x3, x4, x5, x6 over Finite Field of size 2
1
3
B = I.groebner_basis(); B
15
[x1 + 1, x2 + 1, x3, x4 + 1, x5, x6]
```

Conclusion: There is a unique fixed point,

$$(M, A, A_{\text{old}}, B, B_{\text{old}(1)}, B_{\text{old}(2)}) = (x_1, x_2, x_3, x_4, x_5, x_6) = (1, 1, 0, 1, 0, 0)$$

This is exactly what we expected: the *lac* operon is ON.

# Using Sage to compute the fixed points (low lactose)

```
P_{\bullet}<x1,x2,x3,x4,x5,x6> = PolynomialRing(GF(2), 6, order = 'lex'); P
 3
       Multivariate Polynomial Ring in x1, x2, x3, x4, x5, x6 over Finite Field of size 2
    L=0:
    Lh=0:
    print "L =", L;
    print "L h =" . Lh:
       L h = 0
    I = ideal(x1+x2, x2+(L*x4+Lh+x4*L+Lh)+(x2*(1+x3)*(1+x4))+(L*x4+Lh+x4*L*Lh)*(x2*(1+x3)*(1+x4)), x3+(1+x4*L)*(x2*(1+x3)*(1+x4))
    (1+Lh)*x2, x4+x1+x4*(1+x6)+x1*x4*(1+x6), x5+(1+x1)*x4, x6+(1+x1)*x5); I
       Ideal (x1 + x2, x2*x3*x4 + x2*x3 + x2*x4, x2 + x3, x1*x4*x6 + x1*x4 + x1 + x4*x6, x1*x4 + x4 + x5, x1*x5)
12
        + x5 + x6) of Multivariate Polynomial Ring in x1, x2, x3, x4, x5, x6 over Finite Field of size 2
14
    B = I.groebner basis(); B
       [x1 + x6^2, x2 + x6^2, x3 + x6^2, x4 + x6^5 + x6^4 + x6, x5 + x6^4 + x6, x6^6 + x6^4 + x6^3]
```

We need to backsubstitute. Recall that  $x_i^k = x_i$  for all k.

The last equation:  $x_6^6 + x_6^4 + x_6^3 = 0$  implies  $x_6 = 0$ .

Plug this into the previous equation:  $x_5 + x_6^4 + x_6 = 0$  (with  $x_6 = 0$ ) implies  $x_5 = 0$ .

And so on. We get a unique fixed point:

$$(M, A, A_{\text{old}}, B, B_{\text{old}(1)}, B_{\text{old}(2)}) = (x_1, x_2, x_3, x_4, x_5, x_6) = (0, 0, 0, 0, 0, 0)$$

This is exactly what we expected: the *lac* operon is OFF.

# Using Sage to compute the fixed points (medium lactose)

```
P.<x1,x2,x3,x4,x5,x6> = PolynomialRing(GF(2), 6, order = 'lex'); P
3
       Multivariate Polynomial Ring in x1, x2, x3, x4, x5, x6 over Finite Field of size 2
   L=1:
   Lh=0;
   print "L =", L;_
   print "L h =", Lh;
       L = 1
       L h = 0
   I = ideal(x1+x2, x2+(L*x4+Lh+x4*L*Lh)+(x2*(1+x3)*(1+x4))+(L*x4+Lh+x4*L*Lh)*(x2*(1+x3)*(1+x4)), x3+(1+x4*L)*
    (1+Lh)*x2, x4+x1+x4*(1+x6)+x1*x4*(1+x6), x5+(1+x1)*x4, x6+(1+x1)*x5); I
12
       Ideal (x1 + x2, x2*x3*x4^2 + x2*x3 + x2*x4^2 + x4, x2*x4 + x2 + x3, x1*x4*x6 + x1*x4 + x1 + x4*x6, x1*x4
       + x4 + x5, x1*x5 + x5 + x6) of Multivariate Polynomial Ring in x1, x2, x3, x4, x5, x6 over Finite Field
        of size 2
   B = I.groebner basis(); B
15
       [x1 + x4 + x6^9 + x6^8 + x6^5 + x6^4, x2 + x4 + x6^9 + x6^8 + x6^5 + x6^4, x3 + x6^9 + x6^5, x4^2 + x4 + x6^6]
       x6^{11} + x6^{10} + x6^{9} + x6^{8} + x6^{6}, x4*x6 + x6^{10} + x6^{9} + x6^{6} + x6^{2}, x5 + x6^{8} + x6^{4}, x6^{12} + x6^{9}
       + x6^5 + x6^4 + x6
```

The last (7th) equation implies  $x_6 = 0$ . The 6th one then implies  $x_5 = 0$ .

The 5th equation gives no information ( $x_4$  can be anything), as does the 4th ( $x_4^2 + x_4 = 0$ ).

The 3rd equation says  $x_3 = 0$ .

The 2nd equation says  $x_2 = x_4$ , and the 1st equation says  $x_1 = x_4$ .

We get two fixed points:

$$(M, A, A_{\mathsf{old}}, B, B_{\mathsf{old}(1)}, B_{\mathsf{old}(2)}) = (x_1, x_2, x_3, x_4, x_5, x_6) = (0, 0, 0, 0, 0, 0), \text{ or } (1, 1, 0, 1, 0, 0).$$

## Fixed points of our model and bistability

Here is a table showing the fixed points of our model, depending on whether extracellular lactose levels are low, medium, or high.

Inducer level	L	L <sub>m</sub>	М	В	$B_{old(1)}$	$B_{old(2)}$	Α	$A_{\text{old}}$	operon
Low lactose	0	0	0	0	0	0	0	0	OFF
High lactose	1	1	1	1	0	0	1	0	ON
Medium lactose	0	1	0	0	0	0	0	0	OFF
Medium lactose	0	1	1	1	0	0	1	0	ON

Suppose lactose concentration is low  $(L=L_m=0)$ , and so the operon is OFF. The current state is

$$(M, A, A_{\mathsf{old}}, B, B_{\mathsf{old}(1)}, B_{\mathsf{old}(2)}) = (x_1, x_2, x_3, x_4, x_5, x_6) = (0, 0, 0, 0, 0, 0),$$

Now, let's change  $L_m$  from 0 to 1, increasing the lactose level to medium. We are now in the 3rd fixed point above, and so the operon is still OFF.

Conversely, suppose lactose concentration is high  $(L = L_m = 1)$ , and so the operon is ON. The current state is

$$(M, A, A_{\text{old}}, B, B_{\text{old}(1)}, B_{\text{old}(2)}) = (x_1, x_2, x_3, x_4, x_5, x_6) = (1, 1, 0, 1, 0, 0),$$

Now, let's change L from 1 to 0, reducing the lactose level to medium. This takes us to the 4th fixed point above, and so the operon is still ON.

# A Boolean model incorporating dilution & degradation, and time-delays

Instead of the a "middle value" (.135 min $^{-1}$ ), let's choose the high estimate  $\gamma_A = .52 \text{ min}^{-1}$ .

This makes the half-life of A (which was  $\widetilde{h_A} = 15.753$ ) much smaller:

$$\widetilde{h_A} = \frac{\ln 2}{\widetilde{\gamma_A}} = 1.260, \qquad \widetilde{h_B} = \frac{\ln 2}{\widetilde{\gamma_B}} = 22.360 \qquad \widetilde{h_M} = \frac{\ln 2}{\widetilde{\gamma_M}} = 1.5$$

In this case, let's choose a much smaller time-step (e.g., t = 1 min).

We can no longer ignore all of the time-delays, so we introduce the following new variables:

- $M_1$ ,  $M_2$  to model the delayed effect (by  $\tau_B = 2$  min) of mRNA on the production of  $\beta$ -galactosidase.
- $A_1$  to model the delayed action of A on the production of mRNA by  $\tau_M = .1$  min.

We will use the following new variables to model dilution & degradation:

- $M_{\text{old}}$  since  $\widetilde{h_M} = 1.5$  is approximately 1 time-step.
- $A_{\text{old}}$  since  $\widetilde{h_A} = 1.26$  is approximately 1 time-step.
- $B_{\text{old}(1)}$ ,  $B_{\text{old}(2)}$  since loss of  $\beta$ -galactosidase is slower.

#### Remark

We really should use more variables, e.g.,  $B_{\text{old}(1)}, B_{\text{old}(2)}, \dots, B_{\text{old}(22)}$  to accurately track the loss of  $\beta$ -galactosidase. However, we will argue shortly why this won't matter.

# A Boolean model incorporating dilution & degradation, and time-delays

## Proposed model

$$\begin{array}{ll} f_{M} = A_{1} \vee (M \wedge \overline{M_{\text{old}}}) & f_{A_{1}} = A \\ f_{M_{1}} = M & f_{A_{\text{old}}} = \left( (\overline{B} \vee \overline{L_{m}}) \wedge \overline{L} \right) \wedge A \\ f_{M_{2}} = M_{1} & f_{B} = M_{2} \vee \left( B \wedge \overline{B_{\text{old}(2)}} \right) \\ f_{M_{\text{old}}} = \overline{A_{1}} \wedge M & f_{B_{\text{old}(1)}} = \overline{M_{2}} \wedge B \\ f_{A} = (B \wedge L_{m}) \vee L \vee (A \wedge \overline{A_{\text{old}}} \wedge \overline{B}) & f_{B_{\text{old}(2)}} = \overline{M_{2}} \wedge B_{\text{old}(1)} \end{array}$$

Analysis of the long-term behavior of this model leads to similar results as the previous one.

Lactose	L	L <sub>m</sub>	М	$M_1$	<i>M</i> <sub>2</sub>	Mold	В	$B_{\text{old}(1)}$	$B_{\text{old}(2)}$	Α	$A_1$	$A_{old}$
Low	0	0	0	0	0	0	0	0	0	0	0	0
High	1	1	1	1	1	0	1	0	0	1	1	0
Medium	0	1	0	0	0	0	0	0	0	0	0	0
Medium	0	1	1	1	1	0	1	0	0	1	1	0

### A Boolean version of the 5-variable ODE model

## 5-variable ODE model (Yildirim and Mackey, 2004)

Let M(t)= mRNA,  $B(t)=\beta$ -galactosidase, A(t)= allolactose, P(t)= lac permease, L(t)= lactose (concentrations). Extracellular lactose ( $L_e$ ) is a parameter.

$$\begin{split} \frac{dM}{dt} &= \alpha_M \frac{1 + K_1 (e^{-\mu \tau_M} A_{\tau_M})^n}{K + K_1 (e^{-\mu \tau_M} A_{\tau_M})^n} + \Gamma_0 - \widetilde{\gamma_M} M \\ \frac{dB}{dt} &= \alpha_B e^{-\mu \tau_B} M_{\tau_B} - \widetilde{\gamma_B} B \\ \frac{dA}{dt} &= \alpha_A B \frac{L}{K_L + L} - \beta_A B \frac{A}{K_A + A} - \widetilde{\gamma_A} A \\ \frac{dP}{dt} &= \alpha_P e^{-\mu (\tau_B + \tau_P)} M_{\tau_B + \tau_P} - \widetilde{\gamma_P} P \\ \frac{dL}{dt} &= \alpha_L P \frac{L_e}{K_{L_e} + L_e} - \beta_{L_e} P \frac{L}{K_{L_e} + L} - \alpha_A B \frac{L}{K_L + L} - \widetilde{\gamma_L} L \end{split}$$

We'll use the same estimates for degradation and delay constants as in the 3-variable model:

$$\mu=.03~\mathrm{min}^{-1}, \qquad \widetilde{\gamma_A}=\gamma+\mu=.044, \qquad \widetilde{\gamma_B}=\gamma+\mu=.031, \qquad \widetilde{\gamma_M}=\gamma+\mu=.441.$$

New degradation constants estimated at  $\gamma_L=0.0~{\rm min}^{-1}$ , and  $\gamma_P=.65~{\rm min}^{-1}$ . Delay constant estimate is  $\tau_P=.83~{\rm min}$ .

We need a new parameter to help distinguish high vs. medium extracellular lactose: Lem.

#### A Boolean version of the 5-variable ODE model

#### Model assumptions

- Variables are M, B, A, P, L.
- lacksquare Glucose absent. Extracellular lactose present, two parameters:  $L_e$  and  $L_{em}$ .
- Ignore time-delays (Yildirim and Mackey showed that they do not affect bistability).
- Time-step  $\approx$  12 min.
- Ignore (all  $\ll$  12):  $\tau_M = .10$  min,  $\tau_B = 2$  min,  $\widetilde{h_M} = 1.572$  min.
- Introduce dilution & degradation variables: A<sub>old</sub>, B<sub>old</sub>, L<sub>old</sub>, P<sub>old</sub>.

### Proposed model

$$\begin{split} f_{M} &= A \vee (M \wedge \overline{M_{\text{old}}}) & f_{B} = M \vee \left(B \wedge \overline{B_{\text{old}}}\right) \\ f_{M_{\text{old}}} &= \overline{A} \wedge M & f_{B_{\text{old}}} &= \overline{M} \wedge B \\ f_{A} &= (B \wedge L) \vee (L \wedge L_{e}) \vee \left(A \wedge \overline{A_{\text{old}}} \wedge \overline{B}\right) & f_{P} = M \vee \left(P \wedge \overline{P_{\text{old}}}\right) \\ f_{A_{\text{old}}} &= \left(\overline{B} \vee \overline{L}\right) \wedge \left(\overline{L} \vee \overline{L_{e}}\right) \wedge A & f_{P_{\text{old}}} &= \overline{M} \wedge P \\ f_{L} &= ((P \wedge L_{em}) \vee L_{e}) \vee \left((L \wedge \overline{L_{\text{old}}}) \wedge (\overline{B} \wedge \overline{P})\right) & f_{L_{\text{old}}} &= \left((\overline{P} \vee \overline{L_{em}}) \wedge \overline{L_{e}}\right) \wedge L \end{split}$$

### Justification for $f_A$

$$f_A = (B \wedge L) \vee (L \wedge L_e) \vee \left(A \wedge \overline{A_{old}} \wedge \overline{B}\right)$$

There are 3 ways for allolactose to be available at t + 1:

- (i)  $\beta$ -galactosidase and lactose are present.
- (ii) Internal lactose is present and the concentration of extracellular lacatose is high. This ensures that by time t+1, intracellular lactose concentration is high enough to find available trace amounts of  $\beta$ -galactosidase.
- (iii) The concentration of allolactose is high enough that it won't be reduced below the threshold due to dilution & degradation, or to conversion (by  $\beta$ -galactosidase) to glucose & galctose.

## Justification for $f_L$

$$f_L = ((P \land L_{em}) \lor L_e) \lor \left((L \land \overline{L_{old}}) \land (\overline{B} \land \overline{P})\right)$$

There are 3 ways for intracellular lactose to be available at t + 1:

- (i) Lac permease and extracellular lactose are available.
- (ii) There are high levels of extracellular lactose available (even if *lac* permease level is low).
- (iii) There is enough lactose in the cell that it won't be lost to dilution & degradaton, transport out, or conversion into allolactose (by  $\beta$ -galactosidase).

#### Model:

$$\begin{split} f_{M} &= A \vee (M \wedge \overline{M_{\text{old}}}) & f_{B} = M \vee \left(B \wedge \overline{B_{\text{old}}}\right) \\ f_{M_{\text{old}}} &= \overline{A} \wedge M & f_{B_{\text{old}}} &= \overline{M} \wedge B \\ f_{A} &= (B \wedge L) \vee (L \wedge L_{e}) \vee \left(A \wedge \overline{A_{\text{old}}} \wedge \overline{B}\right) & f_{P} = M \vee \left(P \wedge \overline{P_{\text{old}}}\right) \\ f_{A_{\text{old}}} &= \left(\overline{B} \vee \overline{L}\right) \wedge \left(\overline{L} \vee \overline{L_{e}}\right) \wedge A & f_{P_{\text{old}}} &= \overline{M} \wedge P \\ f_{L} &= ((P \wedge L_{em}) \vee L_{e}) \vee \left((L \wedge \overline{L_{\text{old}}}) \wedge (\overline{B} \wedge \overline{P})\right) & f_{L_{\text{old}}} &= \left((\overline{P} \vee \overline{L_{em}}) \wedge \overline{L_{e}}\right) \wedge L \end{split}$$

#### Fixed points:

Ext. Lactose	Le	Lem	М	$M_{\rm old}$	В	$B_{old}$	Α	$A_{old}$	L	$L_{\text{old}}$	P	Pold
Low	0	0	0	0	0	0	0	0	0	0	0	0
High	1	1	1	0	1	0	1	0	1	0	1	0
Medium	0	1	0	0	0	0	0	0	0	0	0	0
Medium	0	1	1	0	1	0	1	0	1	0	1	0